

· 其他肝病 ·

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胸腺基质淋巴细胞生成素在对乙酰氨基酚诱导的急性肝损伤小鼠模型中的作用机制

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摘要: 目的 探讨胸腺基质淋巴细胞生成素(TSLP)在对乙酰氨基酚(APAP)诱导的急性肝损伤小鼠模型中的作用及其机制。方法 16只野生型(WT)雄性C57BL/6J小鼠被随机分为2组,分别为control组和APAP组,每组8只;APAP组按照400 mg/kg的剂量腹腔注射APAP溶液建模,control组注射等体积生理盐水,6 h后进行取材。全自动化学分析仪检测血清ALT及AST,实时定量PCR方法检测肝组织炎症因子TNF- α 和IL-6的mRNA表达,试剂盒检测肝组织匀浆中谷胱甘肽(GSH)含量,实时定量PCR、Western Blot方法检测TSLP的转录和蛋白水平的表达。另取22只WT雄性C57BL/6J小鼠,随机分为3组,分别为control组($n=8$)、APAP组($n=8$)和APAP+rTSLP组($n=6$),APAP+rTSLP组先腹腔注射rTSLP溶液,同时control组、APAP组注射溶剂PBS;30 min后APAP+rTSLP组和APAP组注射APAP溶液,control组注射等体积生理盐水。检测3组小鼠血清ALT及AST;通过HE染色观察小鼠肝脏的病理变化;试剂盒检测肝组织匀浆中氧化应激指标丙二醛(MDA)、超氧化物歧化酶(SOD)水平;Western Blot方法检测自噬相关蛋白LC3 I/II、Beclin1、P62,以及核因子E2相关因子2(Nrf2)、蛋白激酶B(Akt)、磷酸化-Akt(p-Akt)、哺乳动物雷帕霉素靶蛋白(mTOR)、磷酸化-mTOR(p-mTOR)等分子的蛋白表达。此外,取16只WT雄性C57BL/6J小鼠和16只沉默TSLP受体(TSLPR^{-/-})小鼠,分为WT小鼠control组、WT小鼠APAP组、TSLPR^{-/-}小鼠control组和TSLPR^{-/-}小鼠APAP组,每组8只,WT小鼠APAP组和TSLPR^{-/-}小鼠APAP组按照400 mg/kg的剂量腹腔注射APAP溶液建模,WT小鼠control组和TSLPR^{-/-}小鼠control组注射等体积生理盐水。检测4组小鼠血清ALT、AST以及肝组织的MDA含量;Western Blot方法检测LC3 I/II、Akt、p-Akt的蛋白表达。计量资料两组间比较采用成组 t 检验;多组间比较采用单因素方差分析,进一步两两比较采用LSD- t 检验。结果 APAP诱导急性肝损伤小鼠建模成功后,肝脏TSLP的mRNA和蛋白表达水平较control组均升高(P 值均 <0.01)。在应用rTSLP的研究中,相比于control组,APAP组的ALT、AST明显升高(P 值均 <0.001),肝组织HE染色呈现沿中央静脉放射状坏死,氧化应激指标SOD、Nrf2蛋白表达下降,MDA水平上升(P 值均 <0.01);而APAP+rTSLP组较APAP组,ALT、AST下降,肝组织坏死面积减小,SOD、Nrf2蛋白表达升高,MDA下降(P 值均 <0.05);APAP+rTSLP组与control组相比,LC3 I/II、Beclin1、P62、p-Akt、p-mTOR蛋白表达差异均有统计学意义(P 值均 <0.01)。在应用TSLPR^{-/-}小鼠的研究中,建模后,TSLPR^{-/-}小鼠相较于WT小鼠,ALT、AST、MDA升高,LC3 I/II、p-Akt蛋白表达下降(P 值均 <0.01)。结论 TSLP能够增加自噬,降低氧化应激,从而改善过量APAP引起的急性肝损伤,并且其作用机制可能与PI3K/Akt信号通路的激活和mTOR的抑制有关。

关键词: 胸腺基质淋巴细胞生成素; 醋氨酚; 化学性与药物性肝损伤; 小鼠, 近交 C57BL

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Mechanism of action of thymic stromal lymphopoietin in a mouse model of acetaminophen-induced acute liver injury

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Abstract: Objective To investigate the role and mechanism of thymic stromal lymphopoietin (TSLP) in a mouse model of

acetaminophen (APAP)-induced acute liver injury. **Methods** A total of 16 wild-type (WT) male C57BL/6J mice were randomly divided into control group and APAP group, with 8 mice in each group, and the mice in the APAP group were given intraperitoneal injection of APAP solution at a dose of 400 mg/kg to establish an animal model, while those in the control group were given injection of an equal volume of normal saline, with samples collected after 6 hours. An automatic chemical analyzer was used to measure the serum levels of alanine aminotransferase (ALT) and aspartate aminotransferase (AST); quantitative real-time PCR was used to measure the mRNA expression levels of the inflammatory factors tumor necrosis factor- α (TNF- α) and interleukin-6 (IL-6) in liver tissue; the kit was used to measure the content of glutathione (GSH) in liver tissue homogenate; quantitative real-time PCR and Western blot were used to measure the transcriptional level and protein expression level of TSLP. Furthermore, 22 WT male C57BL/6J mice were randomly divided into control group with 8 mice, APAP group with 8 mice, and APAP+recombination TSLP (rTSLP) group with 6 mice; the mice in the APAP+rTSLP group were given intraperitoneal injection of rTSLP solution, while those in the control group and the APAP group were given injection of the solvent PBS; after 30 minutes, the mice in the APAP+rTSLP group and the APAP group were given injection of APAP solution, while those in the control group were given injection of an equal volume of normal saline. The serum levels of ALT and AST were measured; HE staining was used to observe the pathological changes of the liver; kits were used to measure the levels of the oxidative stress indices malondialdehyde (MDA) and superoxide dismutase (SOD) in liver tissue homogenate; Western blot was used to measure the expression levels of the autophagy-related proteins LC3 I/II, Beclin1, and P62 and the molecules such as nuclear factor erythroid 2-related factor 2 (Nrf2), protein kinase B (Akt), phosphorylated Akt (p-Akt), mammalian target of rapamycin (mTOR), and phosphorylated mTOR (p-mTOR). In addition, 16 WT male C57BL/6J mice and 16 TSLP receptor-silenced (TSLPR^{-/-}) mice were divided into WT mouse control group, WT mouse APAP group, TSLPR^{-/-} mouse control group, and TSLPR^{-/-} mouse APAP group, with 8 mice in each group; the mice in the WT mouse APAP group and the TSLPR^{-/-} mouse APAP group were used for modeling by intraperitoneal injection of APAP solution at a dose of 400 mg/kg, and those in the WT mouse control group and the TSLPR^{-/-} mouse control group were given injection of an equal volume of normal saline. The serum levels of ALT and AST and the content of MDA in liver tissue were measured for these four groups, and Western blot was used to measure the protein expression levels of LC3 I/II, Akt, and p-Akt. The independent-samples *t* test was used for comparison of continuous data between two groups; a one-way analysis of variance was used for comparison between multiple groups, and the least significant difference *t*-test was used for further comparison between two groups. **Results** After the mouse model of APAP-induced acute liver injury was established successfully, there were significant increases in the mRNA and protein expression levels of TSLP compared with the control group (both $P<0.01$). In the study of rTSLP, compared with the control group, the APAP group had significant increases in ALT and AST (both $P<0.001$) and radial necrosis along the central vein observed by HE staining of liver tissue, as well as significant reductions in the protein expression levels of the oxidative stress indices SOD and Nrf2 and a significant increase in the level of MDA (all $P<0.01$); compared with the APAP group, the APAP+rTSLP group had significant reductions in ALT and AST, a significant reduction in necrotic area of liver tissue, significant increases in the protein expression levels of SOD and Nrf2, and a significant reduction in MDA (all $P<0.05$); there were significant differences in the protein expression levels of LC3 I/II, Beclin1, P62, p-Akt, and p-mTOR between the APAP+rTSLP group and the control group (all $P<0.01$). In the study of TSLPR^{-/-} mice, compared with the WT mice after modeling, the TSLPR^{-/-} mice had significant increases in the levels of ALT, AST, and MDA and significant reductions in the expression levels of LC3 I/II and p-Akt (all $P<0.05$). **Conclusion** TSLP can increase autophagy, reduce oxidative stress, and thus improve acute liver injury induced by APAP overdose, possibly by activating the PI3K/Akt signaling pathway and inhibiting mTOR.

Key words: Thymic Stromal Lymphopoietin; Acetaminophen; Chemical and Drug Induced Liver Injury; Mice, Inbred C57BL

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对乙酰氨基酚(acetaminophen, APAP)是最常见的非处方解热镇痛药物,然而,其过量使用是导致急性肝损伤和急性肝衰竭最重要的原因,尤其在西方国家^[1-2],且尚无特异的治疗手段。目前对于APAP诱导肝损伤的

发病机制和治疗的认知有限^[3-4],故进一步探究新的分子机制和治疗靶点尤为重要。胸腺基质淋巴细胞生成素(thymic stromal lymphopoietin, TSLP)是一种由4条短链 α 螺旋束组成的I型IL-2家族细胞因子,与IL-7具有同源性,其在免疫调节和促炎调节等方面具有重要作用^[5]。TSLP通过结合由IL-7受体 α 链和TSLP受体(thymic stromal lymphopoietin receptor, TSLPR)链组成的受体复合物促发下游信号^[6]。研究^[7]表明,TSLP在肝脏疾病中具有重要作用,包括良性肝病、肝肿瘤等。TSLP通过PI3K/Akt信号通路保护肝脏缺血再灌注损伤,其机制是TSLP促进自噬的激活^[8];自噬在APAP诱导的肝损伤中可清除APAP代谢过程中产生的中间毒性产物和受损的细胞器,从而起到保护作用^[9]。然而,TSLP在APAP引起的肝毒性中的作用机制目前尚不明确。因此,本研究旨在探究TSLP在APAP诱导的急性肝损伤中的作用及其机制,希望能够为未来的临床诊疗提供新的靶点。

1 材料与方法

1.1 实验动物 54只野生型(WT)雄性C57BL/6J小鼠分别购于长沙天勤生物公司[动物生产许可证编号:SCXK(湘)2019-0014]和北京维通利华实验动物技术有限公司[动物生产许可证编号:SCXK(京)2016-0006]。16只TSLPR基因全敲除(TSLPR^{-/-})雄性C57BL/6J小鼠购于赛业(苏州)生物科技有限公司[动物生产许可证编号:SCXK(苏)2020-0006],用于实验的小鼠均为纯合子雌、雄小鼠繁殖得到的8~10周龄1代小鼠。全部小鼠被饲养于广西医科大学动物实验中心[动物使用许可证编号:SYXK(桂)2020-0004]。所有小鼠在进行实验操作前需提前一天禁食过夜(16h)。

1.2 主要药品和试剂 APAP购于美国MedChemexpress生物科技有限公司。外源重组小鼠TSLP(recombination mouse TSLP, rTSLP)蛋白购于美国R&D Systems公司(货号555-TS-101)。一抗:TSLP(货号PA5-20321)购于美国Invitrogen公司;GAPDH(货号GB11002)、 β -actin(货号GB11001)和P62(货号GB11239)购于武汉赛维尔生物科技有限公司;LC3(货号WL01506)、Beclin1(货号WL02508)、Akt(货号WL0003b)、p-Akt(磷酸化Akt, Ser473)(货号WLP001a)、哺乳动物雷帕霉素靶蛋白(mammalian target of rapamycin, mTOR)(货号WL02477)、p-mTOR(磷酸化mTOR, Ser2448)(货号WL03694)购于沈阳万类生物科技有限公司;核因子E2相关因子2(nuclear factor-erythroid 2-related factor 2, Nrf2)(货号

D221053)购于上海生工生物工程公司。山羊抗兔二抗(货号ab6702)购于英国Abcam公司。超氧化物歧化酶(superoxide dismutase, SOD)(货号A001-3-2)、谷胱甘肽(glutathione, GSH)(货号A006-2-1)、丙二醛(malondialdehyde, MDA)(货号A003-1-2)检测试剂盒均订购于南京建成生物工程研究所。

1.3 动物分组与处理

1.3.1 验证小鼠肝组织TSLP表达 采用随机数字表法,将16只WT小鼠随机分为control组和APAP组,每组8只。APAP组小鼠腹腔注射以生理盐水为溶剂的APAP溶液,剂量为400 mg/kg;control组小鼠则注射等体积生理盐水。

1.3.2 分析TSLP的功能 实验分为两部分。第一部分:采用随机数字表法,将22只WT小鼠随机分为3组,分别为control组、APAP组和APAP+rTSLP组,其中control组和APAP组各8只小鼠,APAP+rTSLP组6只小鼠。APAP+rTSLP组小鼠先腹腔注射无菌PBS溶解的rTSLP溶液(2 μ g/只),而control组和APAP组注射PBS;30 min后,APAP+rTSLP组和APAP组注射400 mg/kg剂量的APAP溶液,control组小鼠则注射等体积生理盐水。第二部分:采用随机数字表法,将16只WT小鼠和16只TSLPR^{-/-}小鼠分为4组,分别为WT小鼠control组、WT小鼠APAP组、TSLPR^{-/-}小鼠control组和TSLPR^{-/-}小鼠APAP组,每组8只。WT小鼠APAP组和TSLPR^{-/-}小鼠APAP组按照400 mg/kg剂量腹腔注射APAP溶液,WT小鼠control组和TSLPR^{-/-}小鼠control组则注射等体积生理盐水。以上所有小鼠应用APAP或生理盐水6h后,在麻醉下被解剖胸腹腔,用心脏采血法采集血液后收集肝脏。

1.4 血清样本 采集的小鼠血液静置离心后提取血清,在本院检验科通过全自动化学分析仪检测小鼠血清ALT、AST水平。

1.5 肝组织病理学 留取部分新鲜肝组织并用含4%多聚甲醛的固定液浸泡过夜,用于HE染色,经乙醇脱水、二甲苯透明、石蜡包埋、切片(片厚5 μ m)。常规HE染色、中性树胶封片后,光镜下观察肝组织的病理表现。应用Image J软件量化肝组织的坏死面积。

1.6 实时定量PCR 采用Trizol法提取RNA,将所有提取的RNA样品浓度定量在1 000 ng/ μ L,逆转录为cDNA,之后进行扩增反应,选取GAPDH作为内参基因,通过2^{- $\Delta\Delta$ Ct}方法计算相应基因的相对表达水平。相关引物信息见表1。

1.7 Western Blot检测 利用PIRA组织裂解液提取肝

表1 引物序列
Table 1 Primers sequence

引物名称	正向引物(5'-3')	反向引物(5'-3')
GAPDH	GACATGCCGCTGGAGAAAC	AGCCCAGGATGCCCTTTAGT
TSLP	CTCAATCCTATCCCTGGCT	GACTTCTTGTGCCATTCCT
TNF- α	ACGGCATGGATCTCAAAGAC	AGATAGCAAATCGGCTGAGG
IL-6	ACAACCACGGCCTTCCCTA	TCCACGATTTCCCAGAGAACA

组织总蛋白,采用BCA法测定蛋白水平。每个样本分别取50 μ g蛋白,用12%聚丙烯酰胺凝胶电泳,然后转移至PVDF膜。用5%脱脂牛奶室温下封闭PVDF膜1h,然后分别置于相应的一抗溶液中,4 $^{\circ}$ C下孵育过夜。所有抗体稀释浓度均为1:1 000。次日,TBST缓冲液洗膜5次后,将膜置于辣根过氧化物酶结合的二抗中,室温下孵育1h。采用化学发光凝胶成像系统(美国ProteinSimple公司)扫描,然后应用Image J软件评估和量化蛋白条带。

1.8 氧化应激相关指标的检测 按照说明书操作,使用相应的试剂盒检测小鼠肝组织中GSH、MDA、SOD水平。

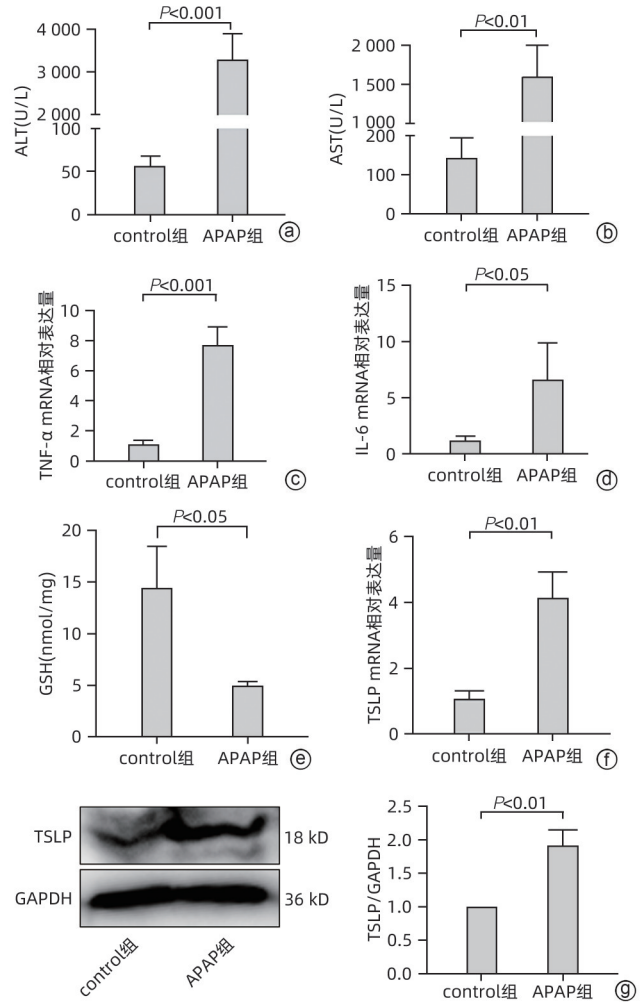
1.9 统计学方法 采用GraphPad Prism 9.0软件进行统计学分析。计量资料以 $\bar{x} \pm s$ 表示,两组间比较采用成组t检验;多组间比较采用单因素方差分析,进一步两两比较采用LSD-t检验。 $P < 0.05$ 为差异有统计学意义。

2 结果

2.1 过量APAP对小鼠肝功能、肝脏炎症及TSLP表达的影响 分组方法如1.3.1节所述。与control组相比,APAP组肝损伤标志物血清ALT、AST和肝组织炎症因子IL-6、TNF- α 的表达明显升高,而肝组织GSH含量明显降低(P 值均 < 0.001);在建模成功的基础上,小鼠肝组织的TSLP蛋白和mRNA表达较control组显著升高(P 值均 < 0.01)(图1)。

2.2 rTSLP对APAP作用小鼠的血清肝酶和肝组织病理的影响 分组方法如1.3.2节第一部分实验所述。APAP组血清ALT、AST较control组升高,APAP+rTSLP组血清ALT、AST较APAP组降低,差异均有统计学意义(P 值均 < 0.05)(图2a、b)。在过量APAP的作用下,小鼠肝组织HE染色表现为沿着中央静脉放射状坏死,而在应用APAP前给予rTSLP能够明显改善上述病理损伤($P < 0.01$)(图2c)。

2.3 rTSLP对小鼠肝脏氧化应激的影响 分组方法如1.3.2节第一部分实验所述。相比于control组,APAP组小鼠肝脏MDA水平升高,SOD和Nrf2水平下降;与APAP组比较,APAP+rTSLP组MDA水平降低,SOD和Nrf2水

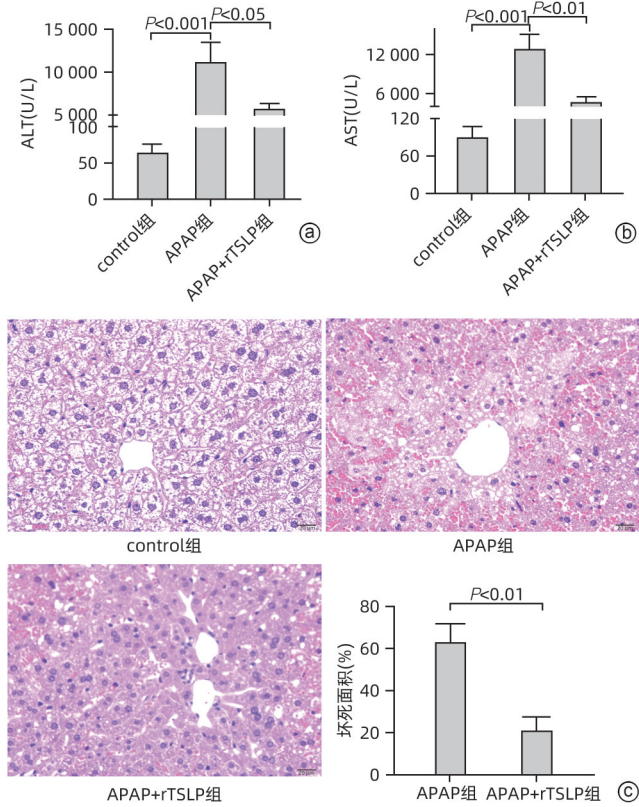


注:a、b分别为两组小鼠血清ALT、AST水平;c、d分别为两组小鼠肝组织炎症因子TNF- α 和IL-6的mRNA表达;e为两组小鼠肝组织GSH水平;f、g分别为两组小鼠肝组织TSLP的mRNA和蛋白表达水平。

图1 血清肝酶、肝组织炎症因子和TSLP表达的比较
Figure 1 The comparison of serum liver enzymes, liver tissue inflammatory factors and TSLP expression of mice in two groups

平上升,差异均有统计学意义(P 值均 < 0.05)(图3)。

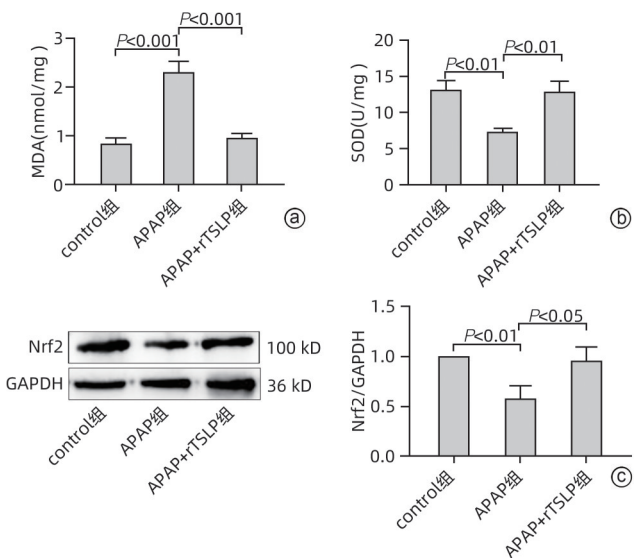
2.4 rTSLP对小鼠自噬发生的影响 分组方法如1.3.2节第一部分实验所述。LC3 I/II、Beclin1、P62是表明自噬发生的重要蛋白。APAP+rTSLP组与control组相比,LC3 I/II、Beclin1、P62的表达差异均有统计学意义(P 值均 < 0.01)(图4)。



注:a,3组小鼠血清ALT水平比较;b,3组小鼠血清AST水平比较;c,3组小鼠肝组织的代表性HE染色结果($\times 200$)及APAP组与APAP+rTSLP组小鼠肝组织坏死面积量化比较。

图2 3组小鼠血清肝酶比较及肝组织学表现

Figure 2 Serum enzymes and hepatic histopathology of mice in the three groups



注:a,3组小鼠肝组织MDA水平比较;b,3组小鼠肝组织SOD水平比较;c,3组小鼠肝组织Nrf2蛋白表达水平及其量化比较。

图3 3组小鼠肝组织氧化应激水平

Figure 3 Oxidative stress levels in liver tissues of mice in the three groups

2.5 rTSLP对小鼠肝脏PI3K/Akt信号通路和mTOR信号通路活化的影响 分组方法如1.3.2节第一部分实验所述。APAP+rTSLP组与control组相比,p-Akt/Akt和p-mTOR/mTOR差异均有统计学意义(P 值均 < 0.01) (图5)。

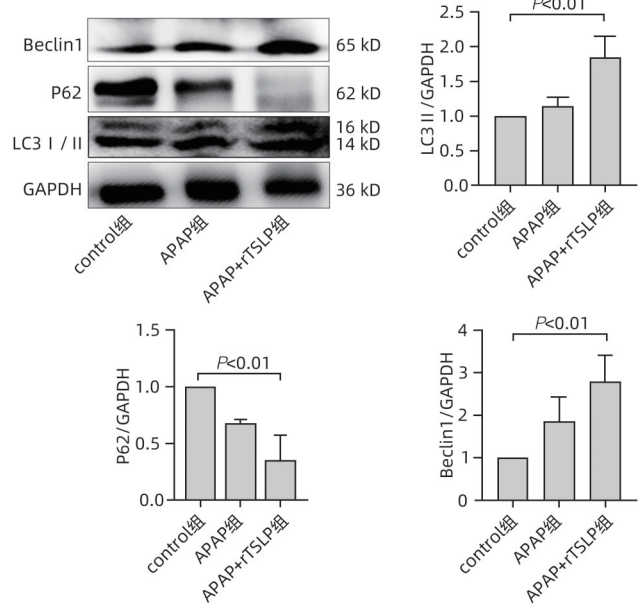


图4 3组小鼠肝组织自噬相关蛋白LC3 I/II、Beclin1及P62的表达

Figure 4 The level of autophagy-related proteins LC3 I/II, Beclin1 and P62 in liver tissues of the three groups

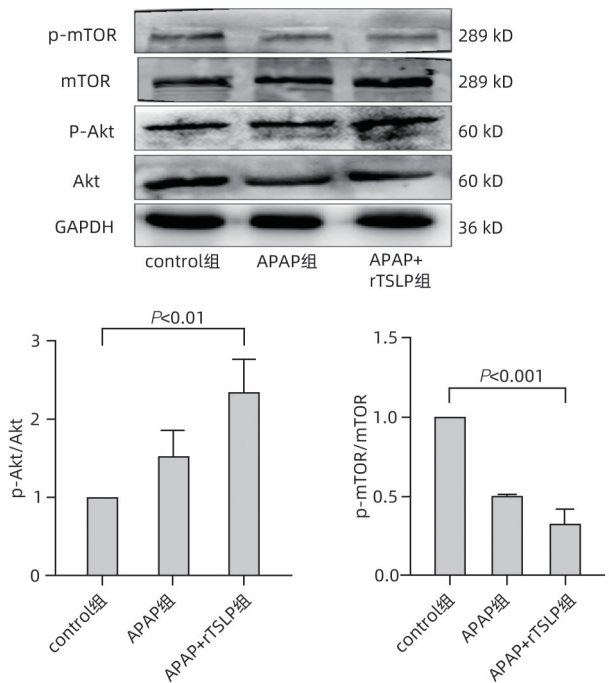


图5 3组小鼠Akt、p-Akt、mTOR及p-mTOR的蛋白表达
Figure 5 Protein expressions of Akt, p-Akt, mTOR and p-mTOR in mice liver of three groups

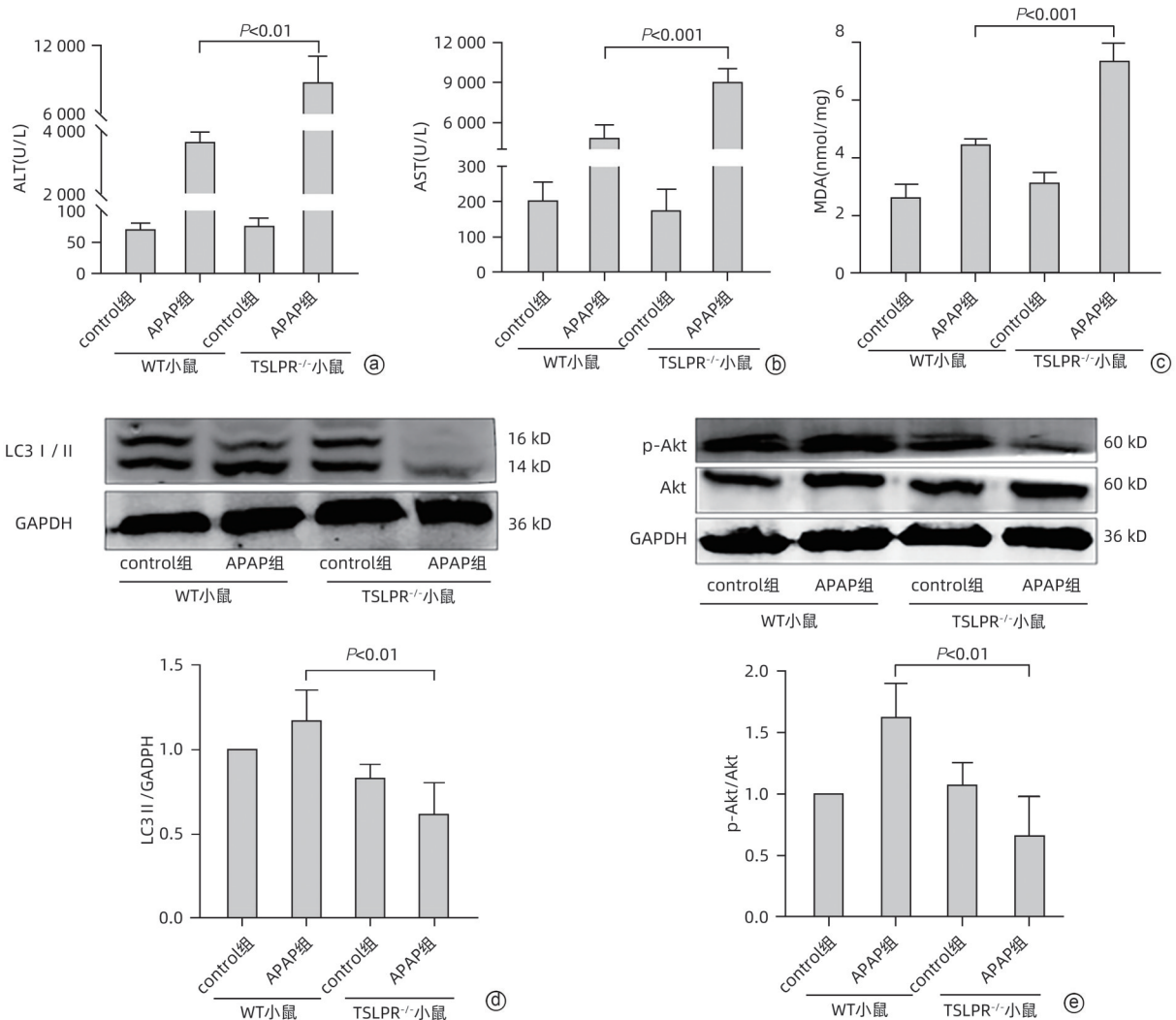
2.6 TSLPR^{-/-}小鼠与WT小鼠建模后的比较 分组方法如1.3.2节第二部分实验所述。相比于WT小鼠APAP组, TSLPR^{-/-}小鼠APAP组ALT、AST、MDA水平均明显升高,自噬蛋白LC3 I/II表达显著降低(*P*值均<0.01);就PI3K/Akt信号通路而言, Akt活化程度更低(*P*<0.01)(图6)。

3 讨论

TSLP主要由上皮细胞分泌,其在炎症、自身免疫性疾病、肿瘤中具有非常重要的作用。虽然TSLP是过敏性因子,但有研究^[10]发现,TSLP在许多非过敏性疾病的病理生理过程中亦发挥了作用。

TSLP在肝脏中表达丰富,尤其是肝细胞,在HCV感染导致慢性丙型肝炎伴冷球蛋白血症患者的肝脏中,

TSLP由肝细胞和角质细胞分泌,并且其促进疾病的进展^[11]。Li等^[8]对TSLP在肝脏缺血再灌注损伤中的作用进行了研究,初步检测到rTSLP的应用可能会降低APAP诱导急性肝损伤小鼠的血清ALT水平,但其作用机制尚未得知。刀豆球蛋白A(Concanavalin A, ConA)也是一种常见的导致肝损伤的药物,在其诱导的肝损伤中, TSLP/TSLPR信号可加重肝毒性^[12]。ConA与APAP诱导的肝损伤均为药物诱导的肝脏无菌性炎症,但TSLP在两者中所起的作用却截然相反,为此,本研究对TSLP在APAP引起的肝毒性中的作用予以进一步探讨。在ConA诱导的肝损伤中, TSLP和IL-4在肝脏可能相互作用形成前馈的炎症级联反应,从而促进嗜酸性粒细胞在肝脏的分布,并且TSLP通过作用于不同亚型的粒细胞



注:a,血清ALT水平比较;b,血清AST水平比较;c,肝组织MDA水平比较;d,肝组织自噬相关蛋白LC3 I/II的表达;e,肝组织Akt和p-Akt蛋白表达。

图6 TSLPR^{-/-}小鼠与WT小鼠的比较
Figure 6 Comparison of TSLPR^{-/-} mice and WT mice after modeling

促进辅助性T淋巴细胞2应答而调节肝损伤^[12]。TSLP在过量APAP诱导的肝损伤小鼠中起到保护作用,主要是因为TSLP对自噬的进一步诱导。氧化应激损伤是APAP肝毒性的中心机制,而自噬在APAP诱导的急性肝损伤中作为保护机制,可清除细胞内受损的细胞器和导致细胞损伤的有害物质,从而改善细胞组织的氧化应激程度,保护肝脏免受APAP引起的氧化应激损伤^[13]。

Nrf2是机体重要的抗氧化保护机制分子,参与药物代谢解毒^[14]。在过量APAP导致的肝损伤中,Nrf2有助于降低APAP代谢过程的高反应性中间代谢物的毒性^[15-16],因此能够活化Nrf2的激动剂,这也许会成为保护肝细胞免受APAP损伤的潜在靶点^[17]。Nrf2为转录因子,其下游包括醌氧化还原酶1、血红素加氧酶1等,因此这些分子组成也被称为Nrf2抗氧化应答元件^[18]。在本研究中,rTSLP蛋白能够提升应用过量APAP注射的小鼠肝组织Nrf2的表达,改善氧化应激程度,因此rTSLP可提升APAP诱导的急性肝损伤小鼠模型的抗氧化能力。

mTOR蛋白是磷酸肌醇3激酶相关激酶家族的一个进化保守的丝氨酸/苏氨酸(Ser/Thr)激酶,其包括两个不同的信号复合物,即mTOR复合物(mTORC)1和mTORC2,mTOR通过调节自噬相关蛋白和溶酶体生物合成,在自噬过程中发挥负调节作用^[19]。本研究结果亦显示,APAP+rTSLP组的自噬发生相较于control组明显增加,而p-mTOR的表达降低。

TSLP能够诱导Akt磷酸化而影响细胞凋亡、增殖、生长和生存。PI3K/Akt信号通路在APAP诱导的肝损伤中是一种保护性的信号通路^[20]。有研究^[8]证明,TSLP能磷酸化活化Akt,与自噬的诱导相关。本研究显示,在过量APAP诱导的肝毒性中,就PI3K/Akt信号通路而言,腹腔注射rTSLP蛋白的小鼠肝组织自噬发生和p-Akt表达较APAP小鼠有升高趋势;沉默了TSLPR基因的小鼠相较于WT小鼠而言,在建模后肝组织的p-Akt表达下调,此时,表征自噬发生的蛋白LC3 I/II水平亦降低。

然而,PI3K/Akt信号通路和mTOR信号通路在APAP肝损伤中的关系还有待进一步验证。已有多项研究^[21-22]表明,PI3K/Akt信号通路活化时能够促进mTOR的磷酸化,PI3K/Akt信号磷酸化增强时,能够使mTOR的磷酸化水平升高,自噬发生水平下降,而本研究表明,APAP+rTSLP组小鼠肝脏中自噬发生较control组增加,同时p-Akt表达升高,而p-mTOR的表达降低,这与之前的研究不一致。因此,在过量APAP诱导的急性肝损伤

小鼠中,PI3K/Akt对于mTOR的作用可能不是主要的,并不能影响自噬的发生,该信号通路可能有其他作用机制,例如增加重要的抗氧化因子Nrf2的表达^[23],其中的机制还有待进一步探究。

综上所述,本研究通过小鼠在体实验表明TSLP能够增加自噬,降低氧化应激,从而改善过量APAP引起的急性肝损伤,并且其作用机制可能与PI3K/Akt信号通路的激活和mTOR的抑制相关,为进一步发现APAP诱导肝损伤的诊疗靶点提供了理论基础,也扩展了关于TSLP在肝脏无菌性炎症领域的认识。

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作者贡献声明: 陈文赏负责课题设计,资料分析,撰写论文;尹明景参与收集数据,修改论文;朱继金负责拟定写作思路,指导撰写文章并最后定稿。

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