

# 支气管肺发育不良发生机制中肺泡上皮细胞转分化关键信号通路的研究进展

张梦玥<sup>1</sup> 周建国<sup>1,2Δ</sup>

<sup>1</sup>国家儿童医学中心/复旦大学附属儿科医院新生儿科 上海 201102;

<sup>2</sup>复旦大学附属儿科医院安徽分院/安徽省儿童医院新生儿科 合肥 230022)

**【摘要】** 支气管肺发育不良(bronchopulmonary dysplasia, BPD)是早产儿严重呼吸系统并发症,重症病例仍缺乏有效治疗手段。BPD是多因素疾病,发病机制主要包括肺泡简单化和肺微血管发育障碍。肺泡上皮细胞是肺泡的主要构成部分,包括肺泡 I 型(alveolar type 1, AT1)和肺泡 II 型(alveolar type 2, AT2)细胞,其中 AT1 细胞参与气屏障构建,发挥气体交换作用,AT2 细胞具有增殖分化的干细胞特性,维持肺内环境稳态、修复肺损伤。肺损伤修复的核心是 AT2 细胞向 AT1 细胞的转分化,而激活转分化的信号转导机制尚未明确。本文通过文献检索和分类总结,探讨肺泡上皮细胞转分化的关键信号转导通路及研究进展,为阐述 BPD 发病机制及探索 BPD 新的治疗方案提供参考。

**【关键词】** 肺泡上皮细胞; 转分化; 信号通路; 支气管肺发育不良(BPD)

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## Research progress on key signaling pathways of alveolar epithelial cell transdifferentiation in the pathogenesis of bronchopulmonary dysplasia

ZHANG Meng-yue<sup>1</sup>, ZHOU Jian-guo<sup>1,2Δ</sup>

<sup>1</sup>Department of Neonatology, Children's Hospital, Fudan University/National Children's Medical Center, Shanghai

201102, China; <sup>2</sup>Department of Neonatology, Anhui Provincial Children's Hospital/Anhui Affiliated Children's

Hospital of Fudan University, Hefei 230022, Anhui Province, China)

**【Abstract】** Bronchopulmonary dysplasia (BPD) is a detrimental respiratory complication associated with prematurity that still lacks effective treatment. BPD is a multifactorial disease with a pathogenesis involving alveolar simplification and impaired vascularization. Alveolar epithelial cells are the main components of alveoli including alveolar type I (AT1) and alveolar type II (AT2) epithelial cells. AT1 cells are involved in constructing the air-blood barrier and facilitating gas exchange, while AT2 cells, characterized by proliferative and differentiated stem cell properties, maintain lung homeostasis and contribute to lung injury. The transdifferentiation of AT2 cells into AT1 cells is a core mechanism in the repair of lung injuries, although the key signaling pathway activating transdifferentiation remains unclear. This article introduces the key signaling pathways and research progress in alveolar epithelial cell transdifferentiation through literature retrieval and classification summary, providing a foundation for elucidating the pathogenesis of BPD and exploring new therapeutic regimens for BPD.

**【Key words】** alveolar epithelial cells; transdifferentiation; signaling pathway; bronchopulmonary

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<sup>Δ</sup>Corresponding author E-mail: joezhou@fudan.edu.cn

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dysplasia (BPD)

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肺具有强大的再生和修复功能。肺泡上皮细胞是肺实质主要组成部分,分为肺泡 I 型(alveolar type 1, AT1)细胞和肺泡 II 型(alveolar type 2, AT2)细胞。AT1 细胞又称鳞状肺泡上皮细胞和小肺泡细胞,占肺泡总表面积的 95% 以上,是气血屏障的主要组成部分,参与气体交换<sup>[1]</sup>。AT2 细胞具有增殖和分化功能,参与肺泡形成及损伤后修复,修复过程由具有干细胞性的 AT2 细胞转分化为 AT1 细胞,因此转分化障碍将会导致不良后果<sup>[2-4]</sup>。研究表明肺泡上皮细胞转分化障碍与支气管肺发育不良(bronchopulmonary dysplasia, BPD)<sup>[5]</sup>、慢性阻塞性肺疾病(chronic obstructive pulmonary disease, COPD)<sup>[6]</sup>、特发性肺纤维化(idiopathic pulmonary fibrosis, IPF)<sup>[7]</sup>等多种呼吸系统疾病密切相关。其中 BPD 是早产儿重要并发症,也是当前新生儿诊疗的难点问题,其发病机制、治疗策略亟待进一步深入研究。虽然肺泡上皮细胞转分化障碍及相关信号通路在 BPD 的研究中一直是热门问题,但缺乏 BPD 发生机制中 AT2 细胞转分化为 AT1 细胞的关键信号通路的综述。本文重点聚焦 BPD 转分化过程的关键信号通路,以期为进一步阐述 BPD 发病机制、探索 BPD 新的治疗方案提供理论基础。

**肺泡化障碍在 BPD 发生发展中的作用** 肺泡化是肺泡形成过程。孕晚期胎儿肺发育,肺泡数量增加,肺泡体积减小,肺中隔形成。BPD 是早产儿慢性呼吸系统疾病,由多种因素共同作用导致。BPD 的发生发展主要原因包括肺发育障碍<sup>[8]</sup>、机械通气<sup>[9]</sup>、围产期感染<sup>[10]</sup>、氧化应激反应<sup>[11]</sup>、遗传易感性<sup>[12]</sup>等。肺泡化障碍和肺微血管发育不良是 BPD 的主要病理表现。Coalson 等<sup>[13]</sup>基于新生儿大样本 BPD 病例的肺组织病理学多中心回顾性研究显示, BPD 病例常见的肺部病理学特征包括肺泡数量减少、体积增大、结构简单化、毛细血管畸形、肺泡壁毛细血管纤维化等。Röbler 等<sup>[14]</sup>通过研究高氧环境下兔肺肺泡和微血管变化,发现高氧损伤对肺泡化有十分显著的抑制作用。诸多研究<sup>[15-16]</sup>证实多种损伤因素作用导致的肺泡化障碍是 BPD 发生发展的关键机制。

**肺泡上皮细胞转分化在 BPD 发病机制和肺损**

**伤修复中的作用** BPD 发病机制复杂,高氧暴露肺损伤是 BPD 常用的动物模型。Sun 等<sup>[17]</sup>通过构建高氧新生大鼠模型发现,肺损伤后 AT2 细胞转分化为 AT1 细胞的活动增强,AT2 细胞具有修复小鼠肺损伤和刺激肺血管发育作用。但 AT2 细胞只占肺泡上皮细胞的 16%,肺泡总面积的 5%,维持肺正常气体交换过程的肺泡上皮细胞的 95% 由 AT1 细胞构成。在肺损伤发生后,AT1 细胞数目减少,为维持肺正常生理学功能,AT2 细胞发挥干细胞性增殖作用<sup>[2-4]</sup>,分化为 AT1 细胞,修复肺组织结构和功能<sup>[18]</sup>。但肺泡上皮细胞转分化的机制仍未明确,因此探讨肺泡上皮细胞转分化的关键信号通路,对于揭示 BPD 发病机制、探索 BPD 治疗新方案具有重要意义。

#### 肺泡上皮细胞转分化的关键信号通路

**FGF 信号通路** Brownfield 等<sup>[19]</sup>通过对小鼠肺泡上皮细胞单细胞 RNA 测序分析、标记表达、谱系追踪和克隆分析,首次阐述了成纤维细胞生长因子(fibroblast growth factor, FGF)信号通路在肺泡祖细胞分化及维持 AT2 细胞形态结构中发挥关键作用。AT1 细胞和 AT2 细胞是由一个共同的芽尖祖细胞(bud tip progenitors, BTPs)分化而来<sup>[20]</sup>。发育过程中的 BTPs 和新生 AT2 细胞持续表达 Fgfr2 受体<sup>[21]</sup>。此外,在肺泡分化过程中, Fgfr2 的配体 Fgf7 和 Fgf10 在周围间充质中高表达<sup>[22]</sup>。调节 FGF 信号通路可以促进肺泡细胞增殖、分化,改善肺功能。Bellusci 等<sup>[23]</sup>的研究显示,将 Fgf10 加入小鼠胚胎肺中,远端间充质中的 Fgf10 可诱导肺内胚层扩张、出芽,进而促进小鼠胚胎肺发育。

**TGF $\beta$  和 BMP 信号通路** 利用人肺移植体和体外培养的各类器官模型, Frum 研究团队发现转化生长因子  $\beta$  (transforming growth factor  $\beta$ , TGF $\beta$ ) 信号通路和骨形态发生蛋白(bone morphogenetic protein, BMP)信号通路在 AT2 细胞分化中发挥相反的作用<sup>[24-25]</sup>。在 BTPs 和 RSPO2-阳性间充质细胞之间,表现为抑制 TGF $\beta$  信号通路可以促进 BTPs 向 AT2 细胞分化,而激活 BMP 信号通路则抑制 BTPs 向 AT2 细胞分化。除了以上研究结果, Frum 研究团队<sup>[26]</sup>还发现了在高 Wnt 环境下,抑制

TGF $\beta$ 和激活BMP信号,可观察到体外培养的人肺组织BTPs大量分化成AT2细胞,并分泌AT2细胞板层体。该团队的研究为理解人类肺发育过程中AT2细胞转分化机制提供了新思路。

此外,TGF $\beta$ 与BMP信号通路还是介导药物修复肺损伤和促进肺发育的重要信号通路。如倍他米松、地塞米松等糖皮质激素促进胚胎肺发育的药理机制研究<sup>[27-28]</sup>,发现激素通过抑制肺的TGF $\beta$ 信号通路,促进BTPs分化为AT2细胞,修复肺损伤、刺激胎肺发育。Kobayashi等<sup>[29]</sup>的IPF机制研究,也发现TGF $\beta$ 和BMP信号通路会导致AT2细胞转分化异常与基底细胞错位。

**Wnt信号通路**与IGF-1 Nabhan等<sup>[30]</sup>通过对肺成纤维细胞的单细胞RNA测序、小鼠动物实验、单分子荧光原位杂交,发现Wnt信号通路通过抑制BTPs向AT1细胞分化来维持AT2细胞的干细胞性。大量研究已证实,Wnt信号通路参与细胞增殖、分化和凋亡。而胰岛素样生长因子-1(insulin-like growth factors-1,IGF-1)在肺泡细胞的分化和修复中也起关键作用<sup>[31]</sup>,但IGF-1影响转分化的机制尚不明确。有研究<sup>[32]</sup>显示IGF-1可以直接通过促进AT2细胞增殖和分化,介导肺泡细胞修复和再生。另一些研究<sup>[33]</sup>则认为抑制Wnt信号通路是IGF-1影响AT2细胞分化的重要信号通路。Wnt5a是Wnt家族的一员,本课题组<sup>[34]</sup>前期通过建立高氧诱导的大鼠BPD模型并运用单细胞测序技术,在AT2细胞转分化过程Wnt5a信号通路作用的研究中得出以下结论:Wnt5a的mRNA和蛋白质是AT2转分化的关键因子,具体表现为间充质干细胞来源的细胞外囊泡通过Wnt5a信号通路,抑制AT2细胞转分化为AT1细胞。

**Hippo-YAP信号通路** Hippo-YAP是一种十分稳定的信号通路,对肺组织发育和肺泡干细胞的行为具有重要调控作用,在肺泡上皮细胞的增殖和分化中扮演重要角色<sup>[35]</sup>。具体表现为YAP蛋白维持肺泡上皮细胞内环境稳态、Hippo信号通路通过控制肺泡干细胞活性限制肺器官大小<sup>[36]</sup>。通过研究调控肺泡再生的机制,Digiovanni等<sup>[37]</sup>应用机械张力诱导Hippo信号相关蛋白YAP蛋白激活,促进小鼠肺泡再生。Hippo-YAP信号通路是近年来肺发育和疾病研究的热点。如Mahoney等<sup>[38]</sup>通过研究胎儿肺泡上皮细胞发育过程,发现倘若缺失YAP

蛋白,Sox2的水平和分布将会无法控制,上皮祖细胞也无法正确响应局部TGF $\beta$ 信号通路的调控,最终导致肺发育不良和肺囊性纤维化;并且提示YAP蛋白在肺组织形成的早期阶段具有显著影响Sox2的表达与分化以及调节肺泡上皮细胞的作用。

**Hedgehog信号通路** Hedgehog信号通路与多种肺部疾病的发生和发展密切相关。例如BPD导致肺损伤时,AT2细胞转分化为AT1细胞的启动须通过旁分泌的方式激活Hedgehog信号通路<sup>[39]</sup>。在IPF发生的过程中,Hedgehog信号通路影响肺成纤维细胞增殖、凋亡、迁移、胶原和纤维蛋白生成以及AT2细胞转分化的过程<sup>[40]</sup>。Peng等<sup>[41]</sup>通过特异性删除成年小鼠肺脏中*Shh*基因,靶向抑制Hedgehog信号通路后发现相邻肺间质细胞增殖扩张,而AT2细胞转分化被抑制,提示Hedgehog信号通路在通过维持小鼠远端气道的上皮和间质稳态方面具有重要作用。Hedgehog信号通路还具有促进AT2细胞转分化的作用<sup>[42]</sup>,发挥促进胎儿肺形成、维持肺内环境稳态和促进肺再生的功能。Ingham等<sup>[43]</sup>对Hedgehog信号通路作用的分子机制研究显示,Hedgehog蛋白与Patched 1受体结合,激活Smoothed蛋白,进而引起下游信号转导。

**Notch信号通路** Notch信号通路主要通过4个Notch受体(Notch1~Notch4)和5个配体(Jag1、Jag2、Dll1、Dll3和Dll4)发挥作用<sup>[44]</sup>。Notch信号通路在AT2细胞转分化与肺部疾病的发生发展中扮演重要角色,遗传和单细胞RNA测序数据显示,Notch信号转导在IPF蜂窝状囊性病变中表现活跃<sup>[45]</sup>。Notch信号通路调节肺泡上皮细胞转分化、维持胎儿肺泡和远端细支气管发育和稳态<sup>[46]</sup>。而Wozniak等<sup>[47]</sup>通过整理成人BPD病例预后情况,总结出Notch信号通路和远端气道形态发育不良与早产儿BPD有关。肺损伤后肺泡和小气道的异常修复也会导致成人COPD的发生<sup>[48]</sup>。通过单细胞测序、谱系追踪研究、建立各种肺损伤和小鼠3D类器官模型,Kopan等<sup>[49]</sup>发现Notch信号通路在肺发育过程中可以影响肺泡和气道上皮细胞、促进肺泡干细胞的分化及肺损伤后修复。

**关键信号转导通路之间的相互作用** 通过单细胞测序技术可以发现关键信号通路在不同的细胞类型中呈现不同的组合表达,并以多对混杂的方式与相应配体及下游蛋白相互作用<sup>[50]</sup>。研究<sup>[51]</sup>表

明BMP信号通路通过促进Smad1-Dvl1复合物形成来抑制Wnt信号通路的活性。Notch信号通路可以增强Wnt经典途径 $\beta$ -catenin及其下游的基因表达<sup>[52]</sup>。FGF信号通路通过灭活GSK-3 $\beta$ 来上调Wnt经典途径 $\beta$ -catenin的表达,Wnt信号通路反过来刺激FGF23启动子的活性<sup>[53]</sup>。现有研究中BPD发病机制涉及的信号通路众多,信号通路相互作用的方式有待进一步研究,准确阐明关键信号通路的相互作用网络,有助于我们发现治疗BPD的潜在靶点。

**结语** 支气管肺发育不良作为早产儿重症呼吸系统并发症,主要表现为肺泡化障碍和肺泡简单化,肺泡上皮细胞转分化在肺泡形成及肺损伤修复过程中发挥关键作用,但肺泡上皮细胞转分化的分子机制复杂,涉及信号通路繁多。随着技术的发展,更加精细的单细胞测序技术、建立类器官模型、人工智能、组学分析等已经被运用在肺癌的研究中,并取得了一定的成果。但在BPD领域尚未有此类报道,未来可以结合新技术深入研究参与肺泡AT2细胞转分化的信号通路,探索信号通路下游关键分子,寻求BPD治疗新策略。

**作者贡献声明** 张梦玥 文献检索,论文构思、撰写和修订。周建国 论文构思、撰写和修订。

**利益冲突声明** 所有作者均声明不存在利益冲突。

## 参 考 文 献

- [ 1 ] WEIBEL ER. What makes a good lung? [J]. *Swiss Med Wkly*, 2009, 139(27-28): 375-386.
- [ 2 ] CHAN M, LIU Y. Function of epithelial stem cell in the repair of alveolar injury [J]. *Stem Cell Res Ther*, 2022, 13(1): 170.
- [ 3 ] CHEN Q, LIU Y. Heterogeneous groups of alveolar type II cells in lung homeostasis and repair [J]. *Am J Physiol Cell Physiol*, 2020, 319(6): C991-C996.
- [ 4 ] GANJI F, EBRAHIMI M, SHIRANI A, et al. Epithelial cells/progenitor cells in developing human lower respiratory tract: characterization and transplantation to rat model of pulmonary injury [J]. *Bioimpacts*, 2023, 13(6): 505-520.
- [ 5 ] JING X, JIA S, TENG M, et al. Cellular senescence contributes to the progression of hyperoxic bronchopulmonary dysplasia [J]. *Am J Respir Cell Mol Biol*, 2024, 70(2): 94-109.
- [ 6 ] ZHU W, HAN L, WU Y, et al. Keratin 15 protects against cigarette smoke-induced epithelial mesenchymal transformation by MMP-9 [J]. *Respir Res*, 2023, 24(1): 297.
- [ 7 ] LI Q, WANG Y, JI L, et al. Cellular and molecular mechanisms of fibrosis and resolution in bleomycin-induced pulmonary fibrosis mouse model revealed by spatial transcriptome analysis [J]. *Heliyon*, 2023, 9(12): e22461.
- [ 8 ] GILTMIER AJ, HIGANO NS, WOODS JC, et al. Evaluation of regional lung mass and growth in neonates with bronchopulmonary dysplasia using ultrashort echo time magnetic resonance imaging [J]. *Pediatr Pulmonol*, 2024, 59(1): 55-62.
- [ 9 ] ZHANG EY, BARTMAN CM, PRAKASH YS, et al. Oxygen and mechanical stretch in the developing lung: risk factors for neonatal and pediatric lung disease [J]. *Front Med (Lausanne)*, 2023, 10: 1214108.
- [ 10 ] ABELE AN, TAGLAUER ES, ALMEDA M, et al. Antenatal mesenchymal stromal cell extracellular vesicle treatment preserves lung development in a model of bronchopulmonary dysplasia due to chorioamnionitis [J]. *Am J Physiol Lung Cell Mol Physiol*, 2022, 322(2): L179-L190.
- [ 11 ] BISACCIA P, MAGAROTTO F, D'AGOSTINO S, et al. Extracellular vesicles from mesenchymal umbilical cord cells exert protection against oxidative stress and fibrosis in a rat model of bronchopulmonary dysplasia [J]. *Stem Cells Transl Med*, 2023, 13(1): 43-59.
- [ 12 ] LAVOIE PM, RAYMENT JH. Genetics of bronchopulmonary dysplasia: an update [J]. *Semin Perinatol*, 2023, 47(6): 151811.
- [ 13 ] COALSON JJ. Pathology of bronchopulmonary dysplasia [J]. *Semin Perinatol*, 2006, 30(4): 179-184.
- [ 14 ] GIACOMO R, LABODE J, REGIN Y, et al. Prematurity and hyperoxia have different effects on alveolar and microvascular lung development in the rabbit [J]. *J Histochem Cytochem*, 2023, 71(5): 259-271.
- [ 15 ] APPUHN SV, SIEBERT S, MYTI D, et al. Capillary changes precede disordered alveolarization in a mouse model of bronchopulmonary dysplasia [J]. *Am J Respir Cell Mol Biol*, 2021, 65(1): 81-91.
- [ 16 ] SILVA DM, NARDIELLO C, POZARSKA A, et al. Recent advances in the mechanisms of lung alveolarization and the pathogenesis of bronchopulmonary dysplasia [J]. *Am J Physiol Lung Cell Mol Physiol*, 2015, 309(11): L1239-1272.
- [ 17 ] SUN Y, CHEN C, LIU Y, et al. Adipose stem cells derived exosomes alleviate bronchopulmonary dysplasia

- and regulate autophagy in neonatal rats[J]. *Curr Stem Cell Res Ther*, 2023, 19(6):919-932.
- [18] JOBE AJ. The new BPD: an arrest of lung development [J]. *Pediatr Res*, 1999, 46(6):641-643.
- [19] D-GBROWNFIELD, DE ARCE AD, GHELFI E, *et al.* Alveolar cell fate selection and lifelong maintenance of AT2 cells by FGF signaling [J]. *Nat Commun*, 2022, 13(1):7137.
- [20] MENG X, CUI G, PENG G. Lung development and regeneration: newly defined cell types and progenitor status [J]. *Cell Regen*, 2023, 12(1):5.
- [21] YUAN T, VOLCKAERT T, REDENTE EF, *et al.* FGF10-FGFR2B signaling generates basal cells and drives alveolar epithelial regeneration by bronchial epithelial stem cells after lung injury [J]. *Stem Cell Reports*, 2019, 12(5):1041-1055.
- [22] NIETHAMER TK, STABLER CT, LEACH JP, *et al.* Defining the role of pulmonary endothelial cell heterogeneity in the response to acute lung injury [J]. *Elife*, 2020, 9:e53072.
- [23] BELLUSCI S, GRINDLEY J, EMOTO H, *et al.* Fibroblast growth factor 10 (FGF10) and branching morphogenesis in the embryonic mouse lung [J]. *Development*, 1997, 124(23):4867-4878.
- [24] FRUM T, HSU PP, HEIN RFC, *et al.* Opposing roles for TGF $\beta$ - and BMP-signaling during nascent alveolar differentiation in the developing human lung [J]. *NPJ Regen Med*, 2023, 8(1):48.
- [25] YUAN J, WEGENKA UM, LUTTICKEN C, *et al.* The signalling pathways of interleukin-6 and gamma interferon converge by the activation of different transcription factors which bind to common responsive DNA elements [J]. *Mol Cell Biol* 1994, 14(3):1657-1668.
- [26] NIKOLIĆ MZ, CARITG O, JENG Q, *et al.* Human embryonic lung epithelial tips are multipotent progenitors that can be expanded in vitro as long-term self-renewing organoids [J]. *Elife*, 2017, 6:e26575.
- [27] WALI Z, GOHAR S, WASEEM S, *et al.* Efficacy of antenatal corticosteroid injection in the prevention of neonatal respiratory distress syndrome after C-Section [J]. *PAK J MED SCI*, 2021, 15(8):1874-1876.
- [28] HARIPRIYA PS, LOUIS DM, NAUSHAD N, *et al.* Effect of antenatal betamethasone on respiratory distress syndrome in preterm neonates [J]. *Res J Pharm Technol*, 2022, 4:15.
- [29] KOBAYASHI Y, TATA A, KONKIMALLA A, *et al.* Persistence of a regeneration-associated, transitional alveolar epithelial cell state in pulmonary fibrosis [J]. *Nat Cell Biol*, 2020, 22(8):934-946.
- [30] NABHAN AN, BROWNFIELD DG, HARBURY PB, *et al.* Single-cell Wnt signaling niches maintain stemness of alveolar type 2 cells [J]. *Science*, 2018, 359(6380):1118-1123.
- [31] VOHLEN C, MOHR J, FOMENKO A, *et al.* Dynamic regulation of GH-IGF1 signaling in injury and recovery in hyperoxia-induced neonatal lung injury [J]. *Cells*, 2021, 10(11):2947.
- [32] ZHU Y, LI Y, JIN W, *et al.* Hyperoxia exposure upregulates Dvl-1 and activates Wnt/ $\beta$ -catenin signaling pathway in newborn rat lung [J]. *BMC Mol Cell Biol*, 2023, 24(1):4.
- [33] BAARSMA HA, SKRONSKA-WASEK W, MUTZE K, *et al.* Noncanonical WNT-5A signaling impairs endogenous lung repair in COPD [J]. *J Exp Med*, 2017, 214(1):143-163.
- [34] AI D, SHEN J, SUN J, *et al.* Mesenchymal stem cell-derived extracellular vesicles suppress hyperoxia-induced transdifferentiation of rat alveolar type 2 epithelial cells [J]. *Stem Cells Dev*, 2022, 31(3-4):53-66.
- [35] GOKEY JJ, SNOWBALL J, SRIDHARAN A, *et al.* YAP regulates alveolar epithelial cell differentiation and AGER via NFIB/KLF5/NKX2-1 [J]. *iScience*, 2021, 24(9):102967.
- [36] HOGAN BL, BARKAUSKAS CE, CHAPMAN HA, *et al.* Repair and regeneration of the respiratory system: complexity, plasticity, and mechanisms of lung stem cell function [J]. *Cell Stem Cell*, 2014, 15(2):123-138.
- [37] DIGIOVANNI GT, HAN W, SHERRILL TP, *et al.* Epithelial Yap/Taz are required for functional alveolar regeneration following acute lung injury [J]. *JCI Insight*, 2023, 8(19):e173374.
- [38] MAHONEY JE, MORI M, SZYMANIAK AD, *et al.* The hippo pathway effector Yap controls patterning and differentiation of airway epithelial progenitors [J]. *Dev Cell*, 2014, 30(2):137-50.
- [39] JING J, WU Z, WANG J, *et al.* Hedgehog signaling in tissue homeostasis, cancers, and targeted therapies [J]. *Signal Transduct Target Ther*, 2023, 8(1):315.
- [40] EFFENDI WI, NAGANO T. The Hedgehog signaling pathway in idiopathic pulmonary fibrosis: resurrection time [J]. *Int J Mol Sci*, 2021, 23(1):171.
- [41] PENG T, FRANK DB, KADZIK RS, *et al.* Hedgehog actively maintains adult lung quiescence and regulates repair and regeneration [J]. *Nature*, 2015, 526(7574):578-582.
- [42] GIROUX-LEPRIEUR E, COSTANTINI A, DING VW,

- et al.* Hedgehog signaling in lung cancer: from oncogenesis to cancer treatment resistance [J]. *Int J Mol Sci*, 2018, 19(9):2835.
- [43] INGHAM PW. Hedgehog signaling [J]. *Curr Top Dev Biol*, 2022, 149:1-58.
- [44] TSAO PN, MATSUOKA C, WEI SC, *et al.* Epithelial Notch signaling regulates lung alveolar morphogenesis and airway epithelial integrity [J]. *Proc Natl Acad Sci U S A*, 2016, 113(29):8242-8247.
- [45] BODAS M, SUBRAMANIYAN B, KARMOUTY-QUINTANA H, *et al.* The emerging role of NOTCH3 receptor signaling in human lung diseases [J]. *Expert Rev Mol Med*, 2022, 24:e33.
- [46] LIU X, ZHU X, ZHU G, *et al.* Effects of different ligands in the Notch signaling pathway on the proliferation and transdifferentiation of primary type II alveolar epithelial cells [J]. *Front Pediatr*, 2020, 8:452.
- [47] WOZNAK PS, MAKHOUL L, BOTROS MM. Bronchopulmonary dysplasia in adults: exploring pathogenesis and phenotype [J]. *Pediatr Pulmonol*, 2024, 59(3):540-551.
- [48] WASNICK R, KORFEI M, PISKULAK K, *et al.* Notch1 induces defective epithelial surfactant processing and pulmonary fibrosis [J]. *Am J Respir Crit Care Med*, 2023, 207(3):283-299.
- [49] KOPAN R, ILAGAN MX. The canonical Notch signaling pathway: unfolding the activation mechanism [J]. *Cell*, 2009, 137(2):216-233.
- [50] GRANADOS AA, KANRAR N, ELOWITZ MB. Combinatorial expression motifs in signaling pathways [J]. *Cell Genom*, 2024, 4(1):100463.
- [51] SALAZAR VS, ZARKADIS N, HUANG L, *et al.* Postnatal ablation of osteoblast Smad4 enhances proliferative responses to canonical Wnt signaling through interactions with  $\beta$ -catenin [J]. *J Cell Sci*, 2013, 126(Pt 24):5598-5609.
- [52] LI CT, LIU JX, YU B, *et al.* Notch signaling represses hypoxia-inducible factor-1 $\alpha$ -induced activation of Wnt/ $\beta$ -catenin signaling in osteoblasts under cobalt-mimicked hypoxia [J]. *Mol Med Rep*, 2016, 14(1):689-696.
- [53] FENG J, ZHANG Q, PU F, *et al.* Signalling interaction between  $\beta$ -catenin and other signalling molecules during osteoarthritis development [J]. *Cell Prolif*, 2024, 57(6):e13600.

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## (上接第106页)

- [17] BENNETT PR, BROWN RG, MACINTYRE DA. Vaginal microbiome in preterm rupture of membranes [J]. *Obstet Gynecol Clin North Am*, 2020, 47(4):503-521.
- [18] BAYAR E, BENNETT PR, CHAN D, *et al.* The pregnancy microbiome and preterm birth [J]. *Semin Immunopathol*, 2020, 42(4):487-499.
- [19] SEO SS, AROKIYARAJ S, KIM MK, *et al.* High prevalence of *Leptotrichia amnionii*, *Atopobium vaginae*, *Sneathia sanguinegens*, and factor 1 microbes and association of spontaneous abortion among Korean women [J]. *Biomed Res Int*, 2017, 2017:5435089.
- [20] JACOB-DUBUISSON F, LOCHT C, ANTOINE R. Two-partner secretion in Gram-negative bacteria: a thrifty, specific pathway for large virulence proteins [J]. *Mol Microbiol*, 2001, 40(2):306-313.
- [21] LECUIT M, NELSON DM, SMITH SD, *et al.* Targeting and crossing of the human maternofetal barrier by *Listeria monocytogenes*: role of internalin interaction with trophoblast E-cadherin [J]. *Proc Natl Acad Sci U S A*, 2004, 101(16):6152-6157.

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