

·基础研究·

从海马区胶质细胞CD137L途径探讨电针治疗神经痛的作用机制

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摘要 **目的:**探讨海马CA1区胶质细胞CD137L在电针改善神经痛大鼠机械痛及学习记忆障碍的作用机制。**方法:**将24只SD大鼠按照随机数字表法分为假手术组、模型组和电针组,每组8只。模型组和电针组制备坐骨神经分支选择性损伤(SNI)神经痛模型;假手术组只暴露坐骨神经,但不结扎。造模后第7天开始,电针组对右侧环跳、阳陵泉穴予以电针刺激,频率2 Hz,强度1 mA,每天1次,每次30 min,连续21 d;假手术组和模型组同等条件抓取、固定,但不进行任何干预。SD大鼠造模前及造模后第28天,用Von-Frey测痛仪检测机械痛阈值;造模后28 d采用新物体识别实验检测大鼠学习记忆能力;酶联免疫吸附法检测海马CA1区CD137L、胶质细胞活化标记物(Iba-1、GFAP)和肿瘤坏死因子- α (TNF- α)的浓度;免疫荧光组织化学法检测海马CA1区CD137L及其受体CD137在神经细胞的表达定位情况。**结果:**①与假手术组比较,模型组机械痛阈值和新物体识别指数均显著降低($P < 0.01$);与模型组比较,电针组机械痛阈值和新物体识别指数均显著升高($P < 0.01$)。②与假手术组比较,模型组海马CD137L、Iba-1、GFAP和TNF- α 浓度均显著升高($P < 0.01$),Iba-1和GFAP免疫反应阳性个数明显增多(P 均 < 0.01);与模型组比较,电针组海马CD137L、Iba-1、GFAP和TNF- α 浓度均显著下降(P 均 < 0.05),Iba-1和GFAP免疫反应阳性个数均明显减少($P < 0.01$)。③免疫荧光检测发现,海马CA1区CD137L及其受体CD137与Iba-1、GFAP共表达明显,提示CD137L及其受体CD137主要表达于小胶质细胞、星形胶质细胞,但不表达于神经元。**结论:**电针可能下调海马CA1区胶质细胞CD137L表达,抑制胶质细胞的活化和TNF- α 的释放,减轻神经痛大鼠疼痛及学习记忆障碍。

关键词 神经痛;海马;CD137L;胶质细胞;电针

神经病理性疼痛(简称神经痛)是躯体感觉系统的损害或疾病导致的疼痛,临床表现为痛觉过敏、痛觉超敏等症状,并常伴随记忆障碍^[1-3]。海马作为大脑内侧颞叶的一部分,在伤害性刺激的感受以及疼痛信号的处理过程中发挥重要作用^[4]。神经痛发生时,海马功能呈现出不同寻常的表现,如记

忆缺陷、胶质细胞激活和细胞因子释放等^[5-7]。CA1区作为海马重要的功能分区之一,在痛觉调制和突触可塑性过程中同样扮演着重要角色^[8],疼痛信息可诱导CA1区蛋白表达和细胞功能的改变,如肿瘤坏死因子- α (tumor necrosis factor- α , TNF- α)释放等^[9-11]。

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大量研究证实,电针环跳、阳陵泉穴可显著抑制神经痛,在海马CA1区观察发现,电针可以减少神经痛大鼠疼痛相关神经元电生理活动的异常^[12]。前期研究通过蛋白质组学筛查发现,电针可抑制海马关键蛋白TMEM126A的表达,减轻神经痛敏反应,改善相关学习记忆障碍,而CD137L是TMEM126A的重要结合蛋白^[13-15]。通过文献进一步梳理分析,CD137属于肿瘤坏死因子(tumor necrosis factor, TNF)受体家族的成员,由肿瘤坏死因子受体超家族成员9(tumor necrosis factor receptor superfamily member 9, TNFRSF9)基因编码。CD137是在活化的CD4⁺和CD8⁺ T细胞、NK细胞、DC、巨噬细胞、肥大细胞等细胞上表达的诱导型共刺激受体,其配体为CD137L,表达于巨噬细胞、B细胞等。

在神经痛研究中发现,敲除CD137L基因的小鼠疼痛阈值升高,并且早期使用CD137L抗体能够有效改善痛敏反应;脊髓节段CD137L可能通过调节小胶质细胞的活化和细胞因子的释放,促进神经痛的发生和发展^[16]。然而,目前关于CD137及配体CD137L中枢神经表达及电针镇痛的作用机制不清。基于此,拟探讨海马CD137及CD137L在电针改善神经痛及学习记忆中的作用。

1 实验材料

1.1 实验动物

24只SPF级雄性SD大鼠,体质量160~180g,购自浙江省医学科学院[许可证号:SCXK(浙)2019-0002],饲养于福建中医药大学实验动物中心[许可证号:SYXK(闽)2019-0007]。饲养环境保持适宜的温度、湿度、12h光照明暗交替。本实验操作过程严格按照我国相关实验动物的规定及福建中医药大学动物管理制度,符合实验动物伦理要求(审批号:FJTCMIACUC2021006)。

1.2 仪器设备和试剂

1.2.1 主要仪器设备 Von-Frey测痛仪(美国IITC公司,型号:NC12775);电子针疗仪(型号:SDZ-II)、一次性针灸针(规格:1寸,0.30mm×25mm)均购自于苏州医疗用品厂有限公司;LEICA冰冻切片机(德国徕卡公司,型号:CM1950);LEICA显微镜(德国徕卡公司,型号:DFC425C)。

1.2.2 主要试剂 5-0非吸收性外科缝线(上海浦东金环医疗用品股份有限公司);CD137/CD137L

抗体、小胶质细胞标记物(Ionized calcium-binding adapter molecule 1, Iba-1)抗体、星形胶质细胞标记物(Glial Fibrillary Acidic Protein, GFAP)抗体(美国Thermo试剂公司);GFAP、Iba-1、CD137L、TNF- α 等ELISA试剂盒(武汉伊莱瑞特生物科技股份有限公司)。

2 实验方法

2.1 实验分组

24只SPF级雄性SD大鼠适应性喂养1周后,按照随机数字表法分为假手术组、模型组和电针组,每组8只。

2.2 动物模型制备

参考DECOSTERD的造模方法建立坐骨神经分支选择性损伤(spared nerve injury, SNI)模型^[17]。大鼠经异氟烷麻醉后,剃除皮毛,75%乙醇消毒,切开右后肢皮肤,钝性分离肌肉,充分暴露坐骨神经主干及其远端分支,钝性分离胫神经、腓总神经和腓肠神经,5-0丝线结扎腓总神经和胫神经并剪断,保留腓肠神经,造模后逐层缝合,切口处覆盖青霉素粉末预防感染。假手术组只暴露坐骨神经,不结扎不剪断坐骨神经及分支。

2.3 干预方法

造模后第7天开始,电针组参照《实验针灸学》及前期研究方法^[18]确定大鼠右侧环跳、阳陵泉穴位置,采用常规一次性针灸针(1寸,0.30mm×25mm)分别刺入10mm、5mm,连接电子针疗仪,电针频率为2Hz,强度为1mA,连续波,每天干预1次,每次30min,连续电针21d。假手术组和模型组大鼠同等条件下进行抓取、固定,但不给予电针干预。

2.4 行为学检测

2.4.1 机械痛阈值检测 参考前期研究方法^[18]进行机械痛阈值测试以及数值计算。在造模前及造模后第7天(干预前)、造模后第28天(干预后)检测3组机械痛阈值。每次刺激后待大鼠重新安静后再开始下一次刺激,测量3次得出的平均值记录为测试结果,代入up-and-down方法的公式计算,最终所得数值即为大鼠缩足阈值(paw withdrawal threshold, PWT),以此反映大鼠的机械痛行为学。

2.4.2 新物体识别实验 根据啮齿类动物具备对新物体有探索的偏好本能,在造模后第28天进行新物体识别实验,以反映大鼠的学习记忆能力。

① 适应阶段:将动物依次放入测试箱内适应环境。每只大鼠放置在测试箱内10min任其自由探索,然

后取出放回原笼内休息 10 min,再放入测试箱 10 min,最后归回原笼位。注意每次更换动物测试前必须清除测试箱内残留的粪便和尿液等,并用 75% 乙醇喷洒、擦拭、通风以消除气味。② 训练阶段:准备 2 个完全相同的圆柱体(物体 A 和物体 B),1 个长方体(物体 C)。3 个物体和大鼠的体积匹配并具备一定体质量,以避免在实验过程中因大鼠攀爬而倾倒。先将物体 A 和物体 B 放置在测试箱的左下角和右下角,调试视频分析系统并定义物体和环境的区域范围,打开录像设备。将大鼠背对 2 个物体放入测试箱中。实验人员离开测试房间以避免干扰。大鼠在测试箱内自由探索 10 min 后,取出放回原笼内,间隔 10 min 后进行下一阶段。③ 测试阶段:将左下角的物体 A 取出,换为物体 C。打开录像设备,同样放置大鼠进入测试箱。实验人员离开房间,通过录像观察大鼠探索情况,物体识别测试时间为 10 min。测试完成后将大鼠归回原笼内。通过视频分析系统采集录像,分析记录大鼠相关指标。

2.5 样本采集

造模后第 28 天进行样本采集。用 1% 浓度异氟烷麻醉大鼠后,新鲜快速取脑,分离海马 CA1 区,置入 EP 管液氮冻存待取。灌注取材快速灌入低温生理盐水,缓慢灌入低温 4% 多聚甲醛,取大脑制作冰冻切片。

2.6 酶联免疫吸附法

① 标准品稀释:根据说明书,对标准品进行梯度稀释,混匀备用。② 加样:分别设空白孔、标准品孔、待测样本孔。在酶标包被板上标准品准确加样 50 μL ,待测样本孔中先加样品稀释液 25 μL ,再加待测样本 25 μL ,轻轻晃动混匀。③ 温育:用封板膜封板后置 37 $^{\circ}\text{C}$ 温育 30 min。④ 配液:将 30 倍浓缩洗涤液用蒸馏水 30 倍稀释后备用。⑤ 洗涤:小心揭去封板膜,弃去液体,甩干,每孔加满洗涤液,静置 30 s 后弃去,如此反复 5 次,拍干。⑥ 加酶:加入酶标试剂 50 μL 。⑦ 温育、洗涤:操作同前。⑧ 显色:每孔加入显色剂 A 50 μL ,再加入显色剂 B 50 μL ,轻轻震荡混匀,37 $^{\circ}\text{C}$ 避光显色 10 min。用 450 nm 波长分别测量每个孔的吸光值(A 值)。整个测定的过程应在加入终止液后的 15 min 内完成。通过 Excel 绘制标准曲线可以计算出样品浓度值,并进行统计学分析。

2.7 免疫荧光组织化学

实验结束后取材腰段脊髓包埋,在恒冷冻切片机内进行切片,厚度为 16 μm ,将切好的组织片贴附

在载玻片上;将切片 PBS 洗 5 min \times 3 次,用基因笔画圈,滴加封闭液,置于 37 $^{\circ}\text{C}$ 恒温箱 1 h;甩掉封闭液,分别滴加一抗(小鼠抗 Iba-1、小鼠抗 GFAP、小鼠抗神经元标记物(Neuronal Nuclei, NeuN)、兔抗 CD137、兔抗 CD137L),湿盒 4 $^{\circ}\text{C}$ 过夜;次日, PBS 洗 5 min \times 3 次,再避光加入二抗,置于 37 $^{\circ}\text{C}$ 箱 1 h; PBS 洗 10 min \times 3 次,擦去组织周围的水分后,滴加含 DAPI 的封片剂,盖上载玻片。使用荧光显微镜拍摄每只大鼠海马 CA1 区,每只至少随机选取 5 个视野拍片,并统计分析。

2.8 统计学方法

所有数据经 SPSS 25.0 统计分析。计量资料符合正态分布以($\bar{x} \pm s$)表示,多组间比较采用方差分析。不符合正态分布以中位数和四分位数间距[$M(P_{25}, P_{75})$]表示,多组间比较采用非参数秩和检验。以 $P < 0.05$ 表示差异具有统计学意义。

3 结果

3.1 3 组机械痛阈值、新物体识别能力比较

3 组机械痛阈值采用重复测量方差分析进行统计,数据满足球形检验($P < 0.001$),3 组主体内时间效应差异具有统计学意义($F = 32.54, P < 0.001$),3 组主体间的组别效应差异具有统计学意义($F = 31.92, P < 0.001$),随后进行两两比较。造模后第 7 天,与假手术组比较,模型组和电针组机械痛阈值均显著下降($P < 0.01$)。电针干预 21 d 后,与模型组相比,电针组可显著提高大鼠机械痛阈值($P < 0.01$)。见表 1。3 组新物体识别指数采用单因素方差分析进行统计,造模后第 28 天,与假手术组比较,模型组新物体识别指数显著降低($P < 0.01$),提示大鼠学习记忆能力显著下降;与模型组比较,电针组新物体识别指数显著升高($P < 0.01$),提示大鼠学习记忆能力显著改善。见图 1。

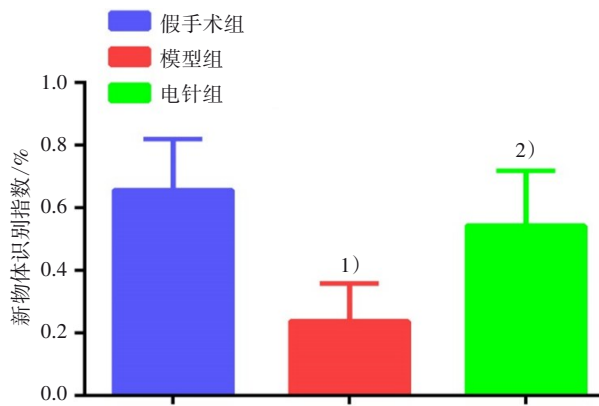
表 1 3 组大鼠机械痛阈值变化($\bar{x} \pm s$)

Table 1 Changes of mechanical pain threshold of rats in three groups ($\bar{x} \pm s$)

组别	<i>n</i>	造模前	造模后第 7 天	造模后第 28 天
假手术组	8	10.13 \pm 1.77	9.50 \pm 1.87	8.78 \pm 2.45
模型组	8	9.51 \pm 1.32	3.34 \pm 1.40 ¹⁾	2.10 \pm 0.43 ¹⁾
电针组	8	9.32 \pm 1.48	3.77 \pm 1.19 ¹⁾	9.80 \pm 2.63 ²⁾

注:与假手术组比较,1) $P < 0.01$;与模型组比较,2) $P < 0.01$ 。

Note: Compared with the sham group, 1) $P < 0.01$; compared with the model group, 2) $P < 0.01$.



注:与假手术组比较,1) $P < 0.01$;与模型组比较,2) $P < 0.01$ 。
 Note: Compared with the sham group, 1) $P < 0.01$; compared with the model group, 2) $P < 0.01$.

图1 3组大鼠新物体识别指数比较 ($\bar{x} \pm s$)
 Figure 1 Comparison of new object recognition in dex in three groups ($\bar{x} \pm s$)

3.2 3组大鼠海马CA1区CD137、CD137L与神经细胞共表达情况

免疫荧光检测表明,海马CA1区CD137免疫反应阳性物质与Iba-1、GFAP免疫反应阳性物质共标记,而与NeuN免疫反应阳性物质未共标记。见图2。同样,在海马CA1区,CD137L免疫反应阳性物质与Iba-1、GFAP共标记,而与NeuN未共标记。见图3。

3.3 3组海马CA1区CD137L浓度的比较

造模后第28天,采用ELISA检测大鼠海马CA1区CD137L浓度。与假手术组比较,模型组CD137L浓度显著上升($P < 0.01$);与模型组比较,电针组CD137L浓度显著下降($P < 0.01$),见图4。

3.4 3组海马CA1区Iba-1、GFAP蛋白表达的比较

造模后第28天,采用免疫荧光和ELISA测定3组海马CA1区小胶质细胞和星形胶质细胞激活标记物的表达。与假手术组比较,模型组Iba-1免疫反应阳性细胞个数和浓度均明显增多($P < 0.01$);与模型组比较,电针组Iba-1细胞个数和浓度均明显减少($P < 0.01$)。见图5和图6。与假手术组比较,模型组GFAP免疫反应阳性细胞个数和浓度均明显增多($P < 0.01$);与模型组比较,电针组GFAP细胞个数和浓度均明显减少($P < 0.01$)。见图7和图8。

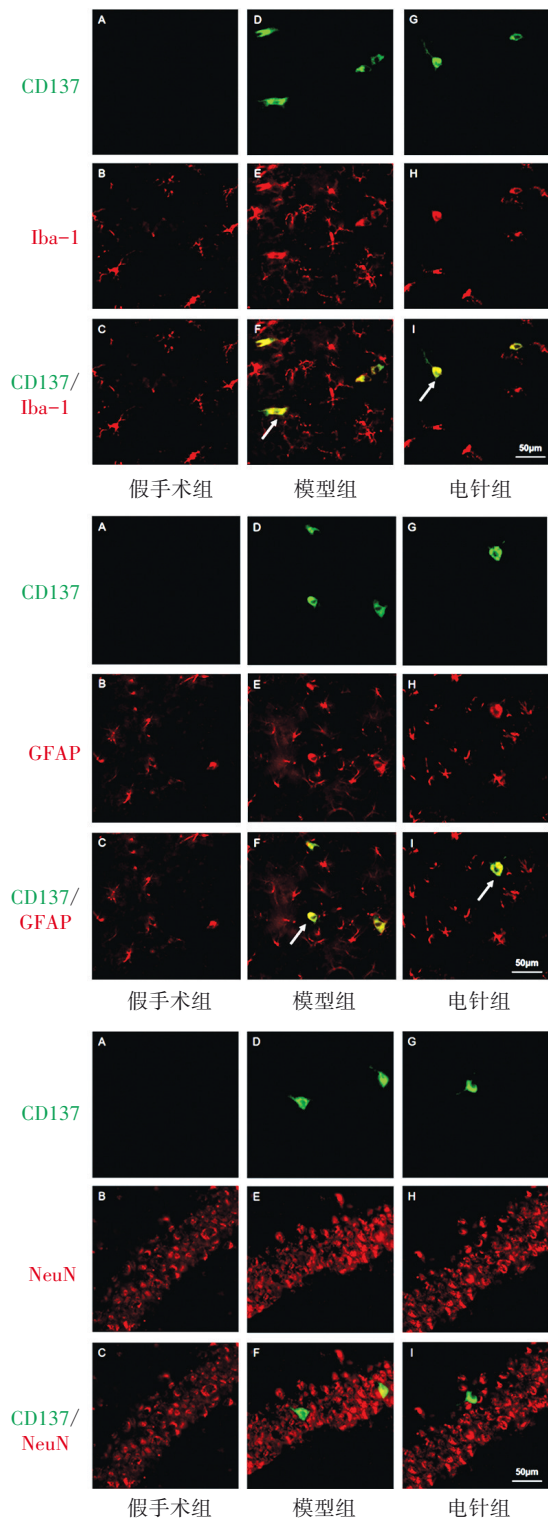


图2 3组海马CA1区CD137和Iba-1、GFAP、NeuN免疫荧光共标情况 ($\times 400$)

Figure 2 Co-labeling of CD137 with Iba-1, GFAP and NeuN immunofluorescence in hippocampal CA1 region of three groups ($\times 400$)

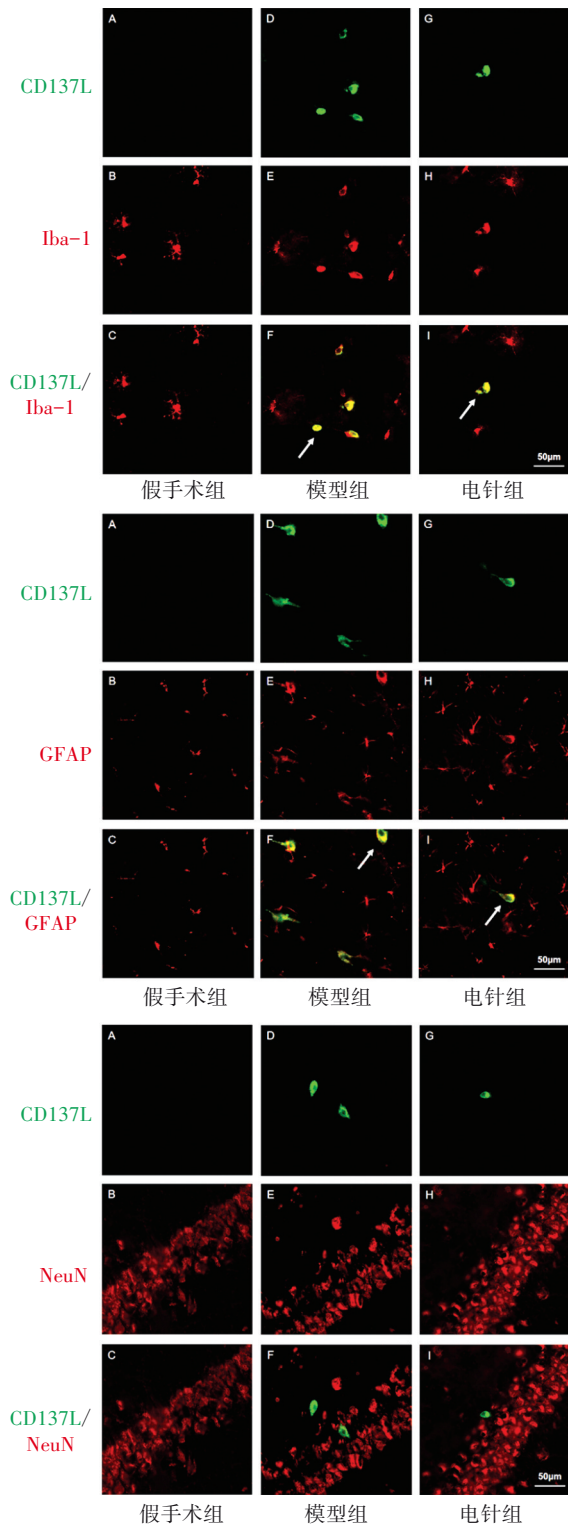
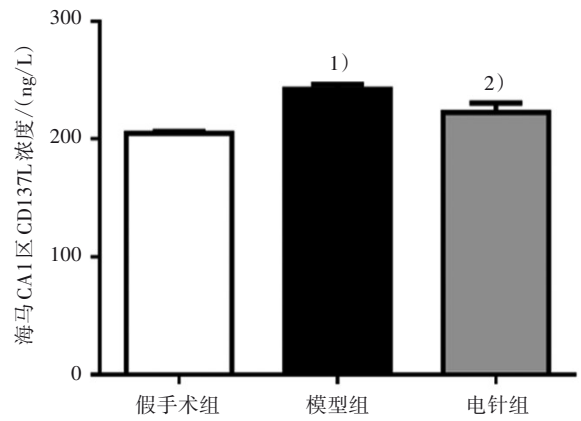


图3 3组海马CA1区CD137L和Iba-1、GFAP、NeuN免疫荧光共标情况(×400)

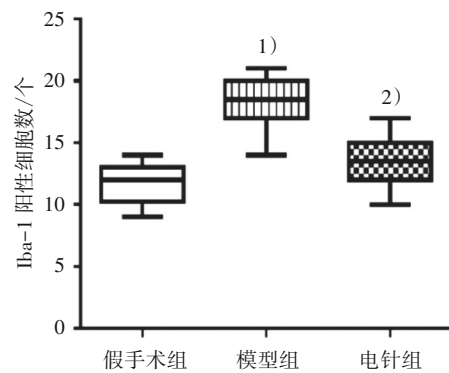
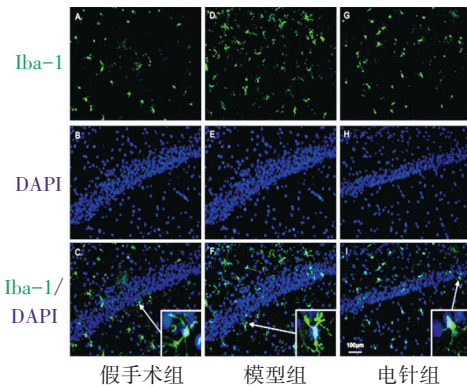
Figure 3 Co-labeling of CD137L with Iba-1,GFAP and NeuN immunofluorescence in hippocampal CA1 region of three groups (×400)



注:与假手术组比较,1) $P<0.01$;与模型组比较,2) $P<0.01$ 。
Note: Compared with the sham group, 1) $P<0.01$; compared with the model group, 2) $P<0.01$.

图4 3组大鼠海马CA1区CD137L浓度比较

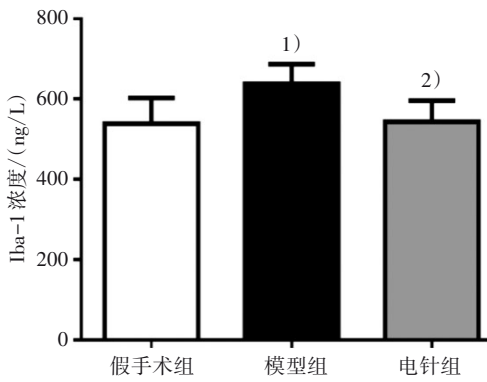
Figure 4 Comparison of CD137L concentration in hippocampal CA1 region in three groups



注:与假手术组比较,1) $P<0.01$;与模型组比较,2) $P<0.01$ 。
Note: Compared with the sham group, 1) $P<0.01$; compared with the model group, 2) $P<0.01$.

图5 3组大鼠海马CA1区小胶质细胞标记物Iba-1表达情况(×200)

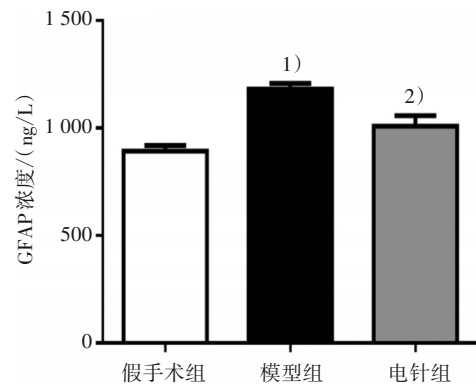
Figure 5 Expression of microglia marker Iba-1 in hippocampal CA1 and the number of immunoreactive cells in three groups (×200)



注:与假手术组比较,1) $P < 0.01$;与模型组比较,2) $P < 0.01$ 。
Note: Compared with the sham group, 1) $P < 0.01$; compared with the model group, 2) $P < 0.01$.

图6 3组大鼠海马CA1区Iba-1含量变化比较

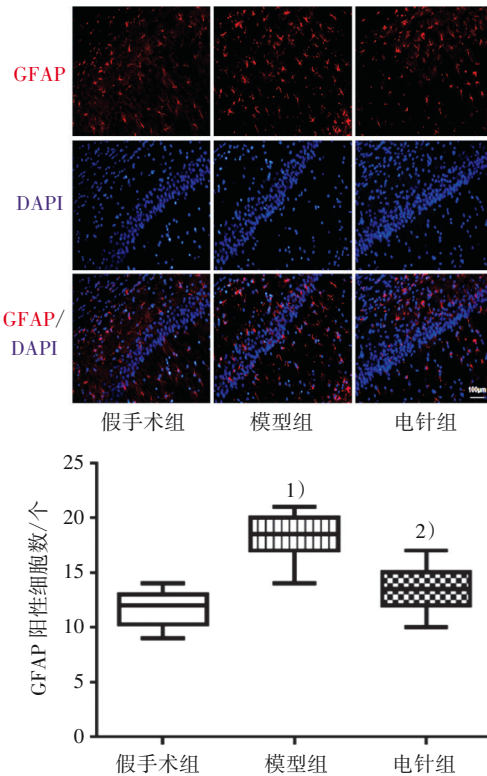
Figure 6 Comparison of Iba-1 content in hippocampal CA1 region of three groups



注:与假手术组比较,1) $P < 0.01$;与模型组比较,2) $P < 0.01$ 。
Note: Compared with the sham group, 1) $P < 0.01$; compared with the model group, 2) $P < 0.01$.

图8 3组大鼠海马CA1区GFAP含量变化比较

Figure 8 Comparison of GFAP content of hippocampal CA1 region in three groups



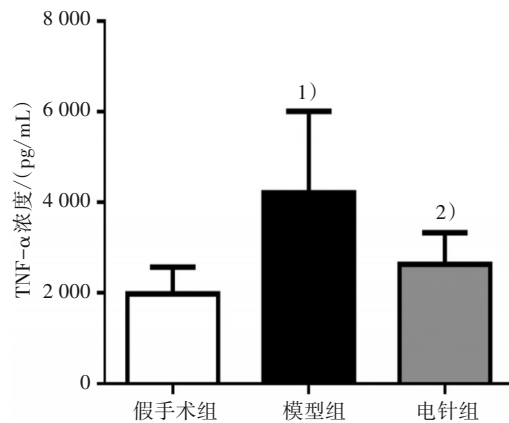
注:与假手术组比较,1) $P < 0.01$;与模型组比较,2) $P < 0.01$ 。
Note: Compared with the sham group, 1) $P < 0.01$; compared with the model group, 2) $P < 0.01$.

图7 3组大鼠海马CA1区星形胶质细胞标记物GFAP表达情况($\times 200$)

Figure 7 Expression of astrocyte marker GFAP in hippocampal CA1 region of three groups ($\times 200$)

3.5 3组大鼠海马CA1区TNF- α 释放浓度比较

造模后第28天,采用ELISA检测大鼠海马CA1区TNF- α 浓度。与假手术组比较,模型组TNF- α 浓度显著上升($P < 0.01$);与模型组比较,电针组TNF- α 浓度显著下降($P < 0.05$)。见图9。



注:与假手术组比较,1) $P < 0.01$;与模型组比较,2) $P < 0.05$ 。
Note: Compared with the sham group, 1) $P < 0.01$; compared with the model group, 2) $P < 0.05$.

图9 3组大鼠海马CA1区TNF- α 浓度比较

Figure 9 Comparison of TNF- α concentration in hippocampal CA1 region of three groups

4 讨论

近年研究表明,海马作为疼痛矩阵脑区的组成之一,是疼痛等伤害性信息加工及修饰的关键脑区,在整合疼痛信息、认知记忆等方面发挥着重要

的作用^[19]。神经痛发生时,海马区长时程增强(long-term potentiation, LTP)减弱,小胶质细胞活化及 TNF- α 释放增多,导致疼痛发生和记忆障碍;抑制海马区胶质细胞活化,降低 TNF- α 释放,可缓解神经痛^[20-22]。

坐骨神经痛归属中医“痛痹证”范畴,应用足少阳经穴环跳、阳陵泉等进行电针,善治诸痛痹不仁,临床康复效果显著。我们前期实验研究同样表明,电针环跳、阳陵泉可显著抑制坐骨神经痛,但关于海马中枢的调控机制仍有待深入明晰^[23-25]。

课题组前期蛋白组学筛选发现,电针可显著提高神经痛海马区 TMEM126A 表达。进一步文献梳理发现, TMEM126A 可能结合 CD137L 结合,在神经痛及电针干预中扮演重要角色^[14]。基于此,本研究将 CD137L 及其受体 CD137 作为电针干预神经痛的可能作用靶点^[26]。实验研究表明,海马 CA1 区 CD137L 及其受体主要表达于胶质细胞,电针可显著抑制神经痛海马区 CD137L 表达和胶质细胞活化,降低炎症因子 TNF- α 的释放。

CD137 又称 4-1BB,是一种有效的 T 细胞共刺激分子,在活化 T 细胞、NK 细胞和血管内皮细胞上表达。其配体 CD137L 是一种跨膜蛋白,在活化的抗原提呈细胞(antigen presenting cell, APC)上与 CD137 结合,并将激活信号传递到 APC 中^[27]。CD137-CD137L 的主要功能是调节细胞存活,抑制二者结合可阻止肿瘤坏死因子受体(tumor necrosis factor receptors, TNFRs)下游信号通路的激活,降低免疫炎症反应^[28-29]。

然而,CD137 和 CD137L 在疼痛疾病中的探索尚属于起步阶段。WAKLEY 等^[16]研究发现,基因敲除 CD137L 的神经痛小鼠表现出显著的痛敏反应,早期鞘内注射 CD137L 抗体提高机械痛阈值,抑制脊髓小胶质细胞炎症反应。体内外实验发现,小胶质细胞与 CD137L 共表达,激活小胶质细胞,诱导炎症因子如 TNF- α 等的释放^[30]。在炎症状态下,CD137L 是巨噬细胞持续产生 TNF- α 所必需的关键蛋白;巨噬细胞表达 CD137L,与 Toll 样受体相互作用,维持 TNF- α 产生^[31]。

综上所述,电针可能是通过海马 CA1 区 CD137L 调节途径,抑制胶质细胞的激活和 TNF- α 的释放,改善神经痛大鼠疼痛及其学习记忆障碍,今后将通过 CD137L 及受体病毒干扰、基因敲除等技术,进一步深入验证 CD137L/CD137 在电针抗炎镇痛中的中枢作用机制。

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Mechanism of Electroacupuncture on Neuropathic Pain via Regulation of CD137L in Hippocampal Glia

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ABSTRACT Objective: To investigate the mechanism of CD137L in CA1 region of hippocampus in improving mechanical pain and memory impairment in neuropathic pain rats during electroacupuncture therapy. **Methods:** A total of 24 SD rats were randomly divided into sham operation group, model group and electroacupuncture group, with 8 rats in each group. In the model and electroacupuncture groups, the spared nerve injury (SNI) model was made. The sham operation group was just exposed right sciatic nerve without ligation. From the 7th day after modeling, the electroacupuncture group received electroacupuncture intervention in the right side Huantiao (GB30) and Yanglingquan (G34) acupoints, 30 minutes each time, once a day, lasting for 21 days. Rats in the model and the sham operation groups were equally grasped and fixed without any intervention. Von-Frey pain detector was used to measure the mechanical withdrawal threshold of rats. On the 28th day after modeling, new object recognition test was used to detect the learning and memory ability. The expression of hippocampal CA1 region CD137L, Iba-1, GFAP and tumor necrosis factor- α (TNF- α) was detected by enzyme-linked immunosorbent assay (ELISA). Immunofluorescence was used for the cellular localization of CD137L and receptor CD137, and to measure the Iba-1 and GFAP expression in hippocampal CA1 region. **Results:** 1) Compared with the sham operation group, mechanical pain threshold and new object recognition index in the model group significantly decreased ($P < 0.01$); compared with the model group, mechanical pain threshold and new object recognition index in the electroacupuncture group significantly increased ($P < 0.01$). 2) Compared with the sham operation group, the concentrations of CD137L, Iba-1, GFAP and TNF- α in hippocampus of rats in the model group significantly increased ($P < 0.01$), and the number of positive Iba-1 and GFAP immune reactions significantly increased ($P < 0.01$); compared with the model group, the concentrations of CD137L, Iba-1, GFAP and TNF- α in hippocampus of rats in the electroacupuncture group significantly decreased (all $P < 0.05$), and the number of positive immune reactions of Iba-1 and GFAP significantly decreased (all $P < 0.01$). 3) Immunofluorescence detection showed that CD137L and its receptor CD137 were significantly co-expressed with microglia marker (Iba-1) and astrocyte marker (GFAP) in hippocampal CA1 region, suggesting that CD137L and CD137 were mainly expressed in microglia and astrocytes, but not in neurons. **Conclusion:** Electroacupuncture may play an analgesic role by reducing the expression of glial CD137L, the amount of activated glial cells and the concentrations of TNF- α .

KEY WORDS neuropathic pain; hippocampus; CD137L; glial cell; electroacupuncture

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Video Kinematic Analysis of Timed Up and Go Test in Convalescent Stroke Patients

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ABSTRACT Objective: This study investigated and compared the results of the Timed Up and Go test (TUG test) between convalescent stroke patients and healthy controls by video recording, and the total time and sub-phase kinematic characteristics of the TUG test were analyzed to provide a theoretical basis for clinical application. Using the convenient sampling method, a total of 41 convalescent stroke patients were recruited as patient group (observation group) in the rehabilitation medical centers of three hospitals (including the First Affiliated Hospital of Nanjing Medical University, etc.) from June to September 2022. Additionally, 35 healthy people with gender, age, height and weight matched were selected as healthy control group (control group). The TUG test were recorded by cameras for all subjects. VirtualDube 2[®] software was used to analyze the kinematic characteristics of recorded video, including the total time and sub-stages parameters (stage 1 standing up, stage 2 forward walking, stage 3 first turning, stage 4 return walking, stage 5 end turning and sitting down); moreover, the observation group also received evaluation including five-times-sit-to-stand test, single leg support time and 10 m maximum walking speed. **Results:** Compared with the control group, the observation group had a significant increase in the time of total TUG [26.91(16.52, 35.98) s] and the sub-stages [1.98 (1.13, 2.48), 8.15 (4.21, 13.02), 4.15 (3.84, 5.93), 6.10 (3.08, 9.94), 5.04 (3.82, 7.03) s] ($P < 0.01$), and there were significant differences in the percentage distribution of each sub-stage ($P < 0.05$). There was a significant positive correlation between five-times-sit-to-stand test and time of total TUG test and each sub-stage in the observation group ($P < 0.01$). There was a significant negative correlation between single leg support time of hemiplegic side and time of total TUG test and each sub-stage ($P < 0.01$). There was a significant positive correlation between 10 m maximum walking speed and time of total TUG test and each sub-stage in the observation group ($P < 0.01$). **Conclusion:** The TUG test video kinematics analysis can be used to accurately analyzing walking characteristics of patients in convalescent stroke patients, and corresponding sub-stage time can reflect the lower limb strength, standing balance, walking stability and other function of stroke patients.

KEY WORDS stroke; Timed Up and Go test; kinematics; video analysis; gait

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